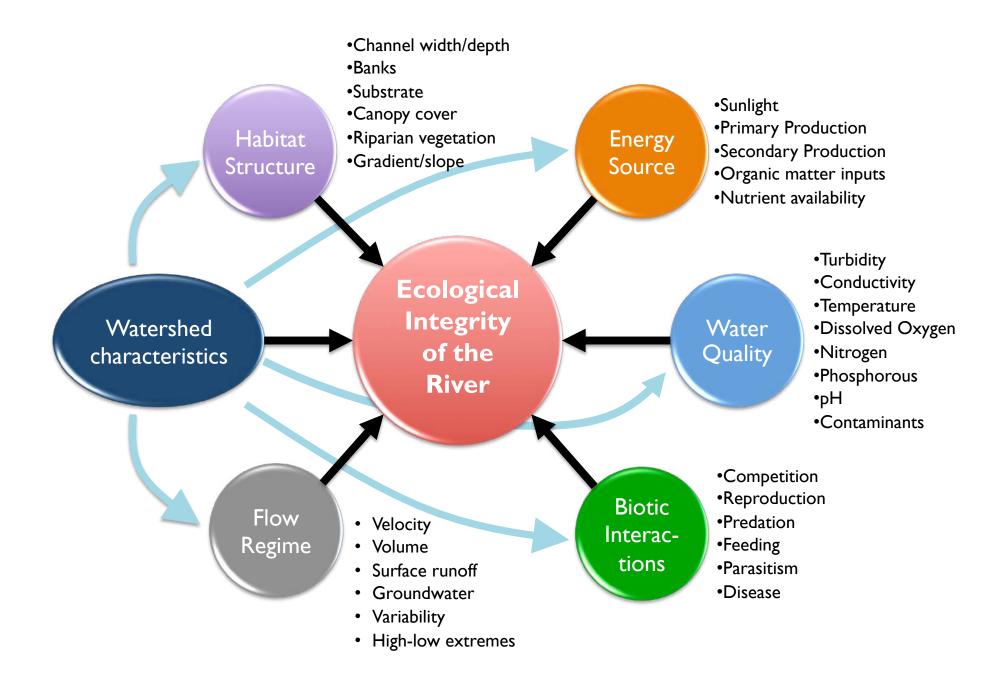
Microorganisms in Freshwaters

Jinjun Kan, PhD Microbiology Stroud Water Research Center





Outline

- Introduction
 - Microorganisms (abundance, mass, type, size, diversity etc.)
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Outline

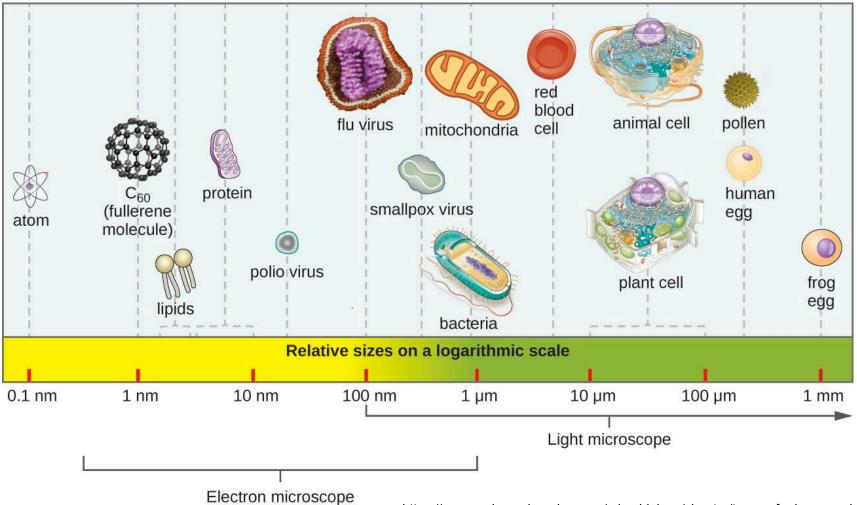
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Micro-organisms

Microscopic; Too small to be seen by unaided eyes

Living organisms, independent



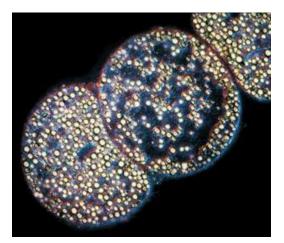
https://courses.lumenlearning.com/microbiology/chapter/types-of-microorganisms/

Different Size

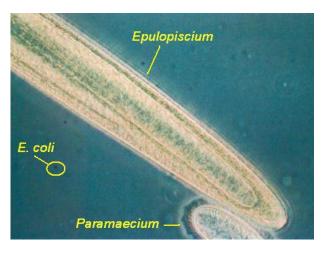
Individual cells: Cocci-most 0.1-0.2 μm in diameter

E. coli/Bacillus: 0.2 μm wide, 1-5 μm long.

A few unusually large cells:



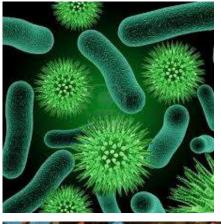
Thiomargarita namibiensis (100-300 µm in diameter)



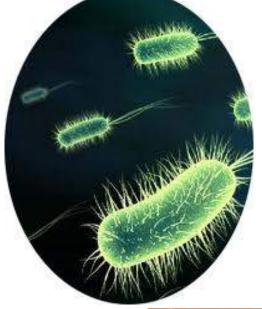
Epulopiscium fishelsoni (80 μm *dia*, 200-700 μm long)

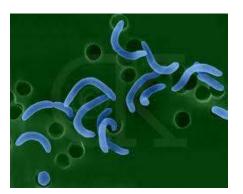
Some form filaments, some in sheaths

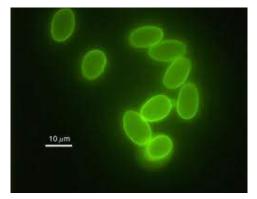
Different type

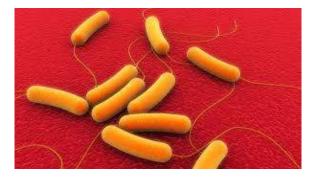


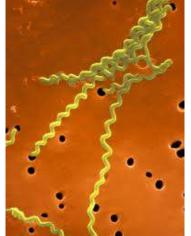






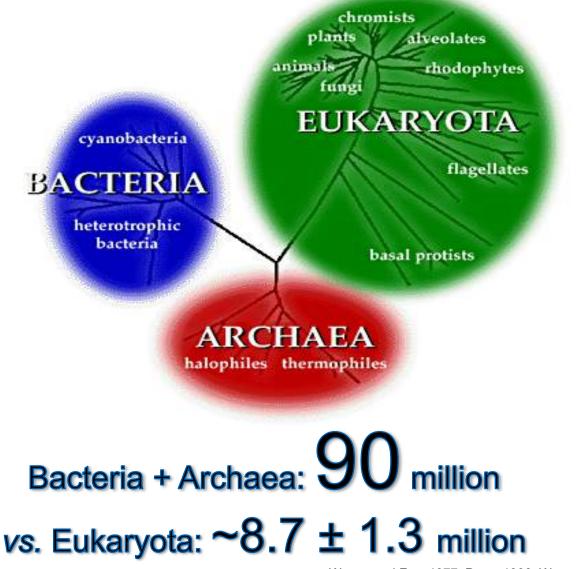






Miscellaneous online resources

High diversity



Woese and Fox, 1977; Pace, 1990; Woese et al., 1990; Mora et al. 2011

Abundance and Mass

Earth is a microbial planet

```
Population size:
```

Human: 7.73 × 10⁹ (estimated Sep 2019)

Microbes: 1.2×10^{31} (including in both water and soils)

 $\sim 1.5 \times 10^{20}$ microbes for every human

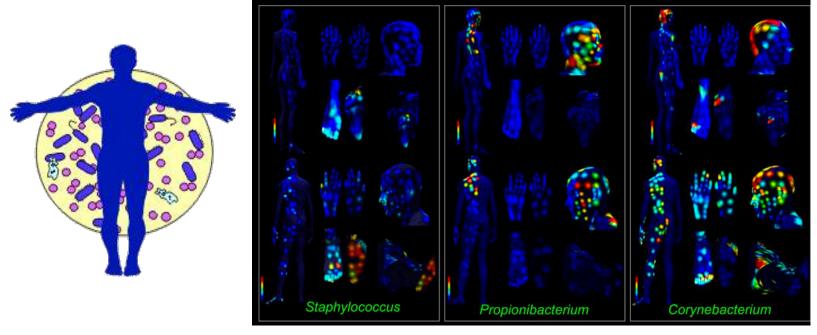
Biomass

```
Humans (@70 kg) = 0.05 Gt C
Microbes = ~ 77.2 Gt C
Microbes "outweigh" humans ~ > 1,500 to 1
```

Microbes are second largest pool of living C (after plants) and the largest pool of living N and P

Human microbiome: a good analogy

Human is a microbial "body"



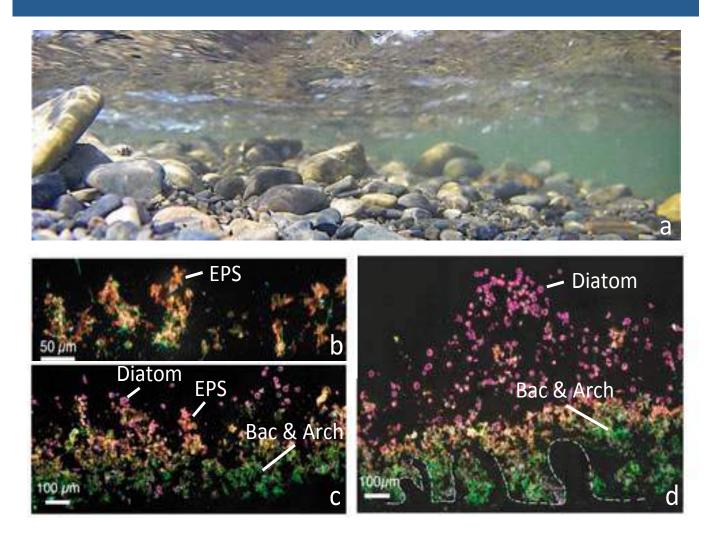
Bouslimani et al. 2015

Cells in human body: 37.2 trillion; Bacteria: 1-10 times more

Outline

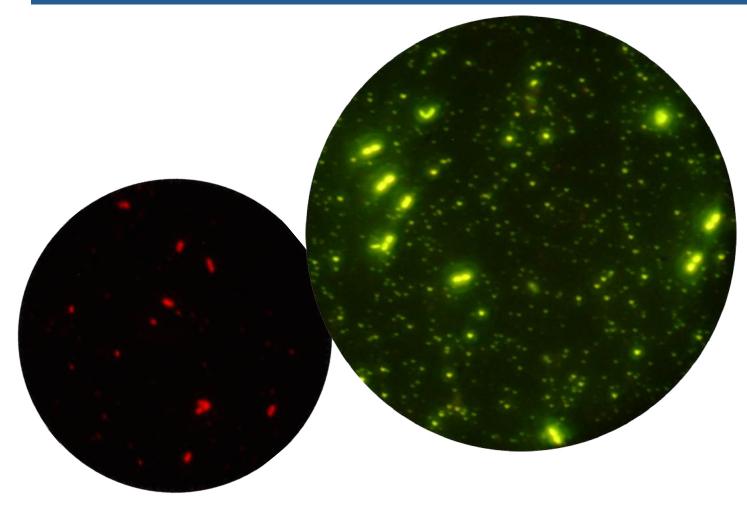
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Microbes living in freshwater



Planktonic (free-floating) vs. Benthic (biofilm); 1 million cells/ml water

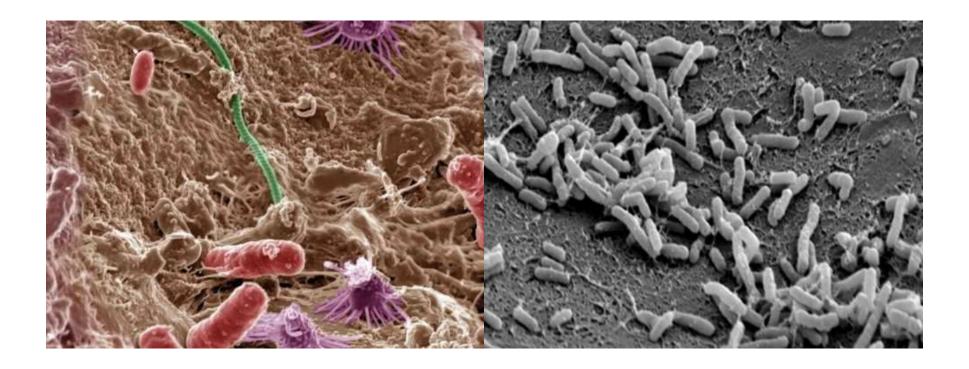
Under microscope



Photosynthetic pigments

Epifluorescence (DNA staining)

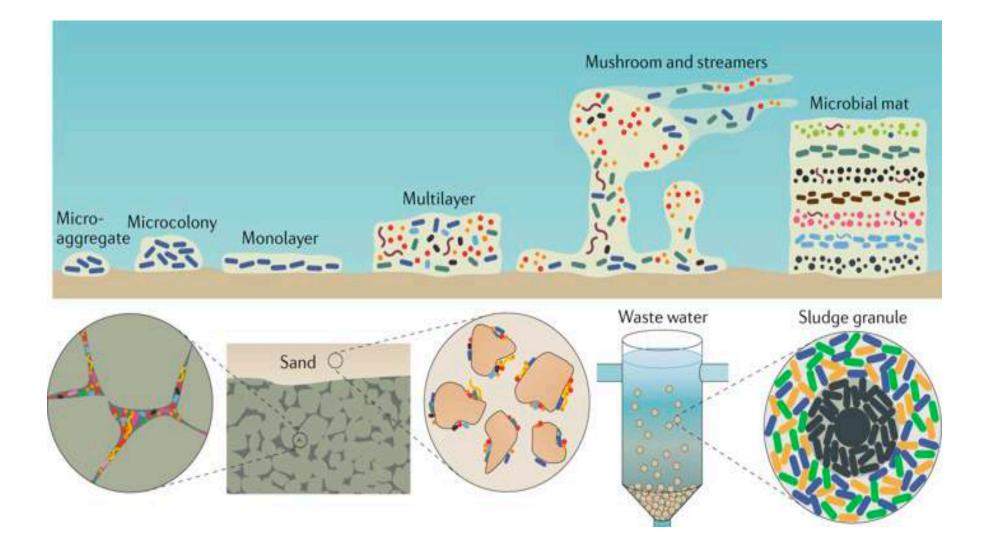
Microbes living in soil/sediments

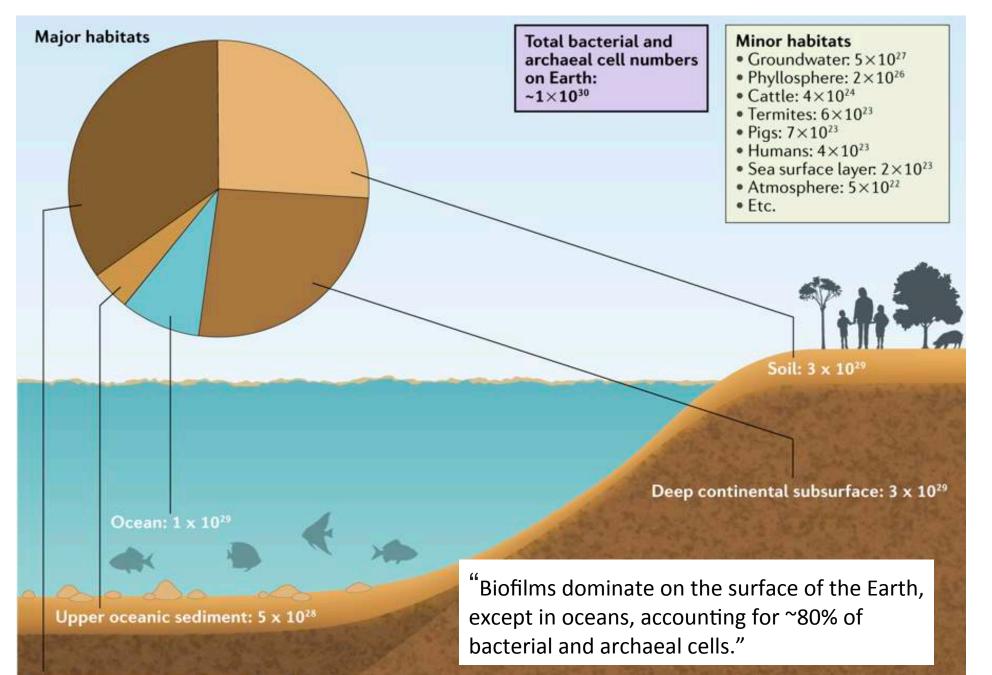


Microbes growing in soils; up to 1 billion cells/g dry soil

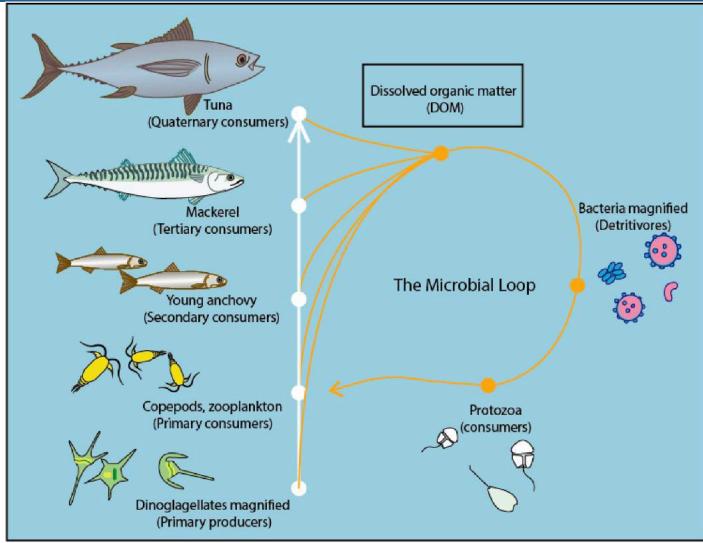
https://www.pitchcare.com/news-media/functioning-healthy-soil.html

Biofilms: "microbial skin"





Significance in ecosystems

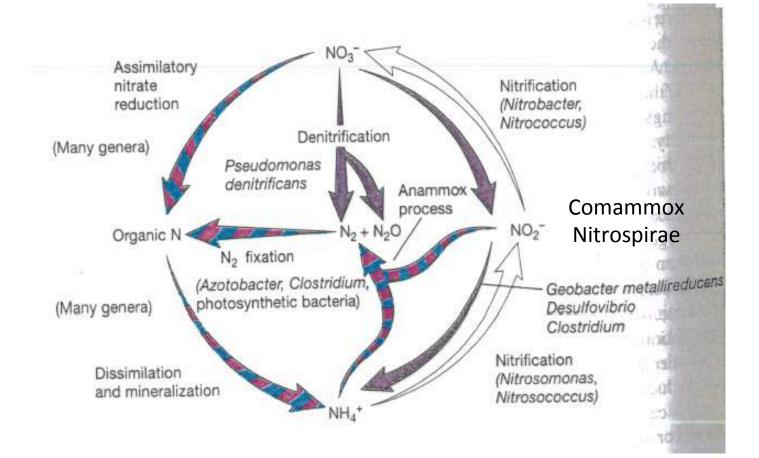


Food web and transformation of dissolved organic matter (microbial loop)

http://www.plankton-bloom.com/ocean-processes/marine-snow/

Nutrient cycling

Significance: Transformation of key elements including Carbon, Nitrogen, Phosphorus, Sulfur, etc.



Public Health Concerns

Water-representative taxa and problems

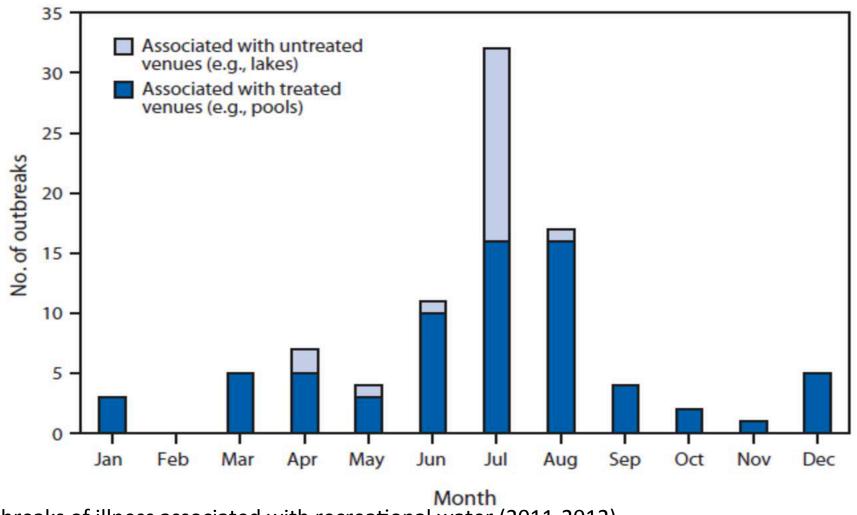
- Pseudomonas aeruginosa: ear infection, bathing beaches
- Clostridium botulinum: food poisoning
- Legionella pneumophilia: respiratory infection and death Legionaire's disease
- Salmonella, Shigella, Vibrio et al.:
 - gi (gastrointestinal) tract, diarrhea and dysentery
- Streptococcus, Vibrio vulnificus et al.:

Necrotizing fasciitis

Cryptosporidium, Giardia etc.

Public Health Risk

- May cause a variety of diseases



Outbreaks of illness associated with recreational water (2011-2012)

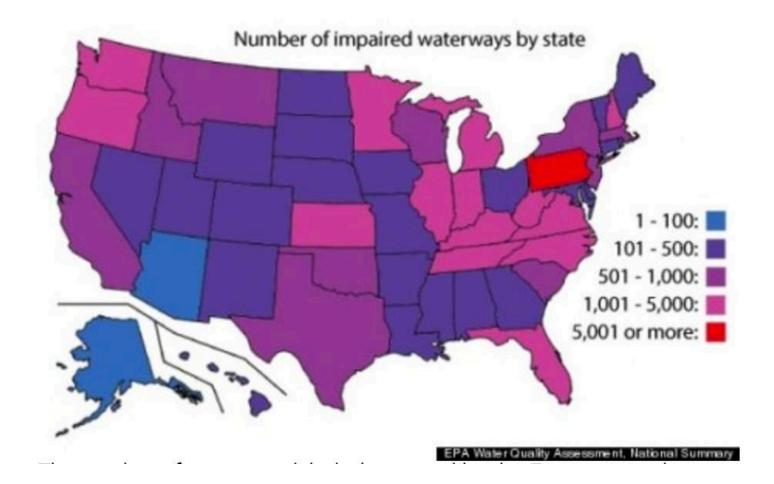
Hlavsa et al. 2015 CDC

Bacterial monitoring for water quality

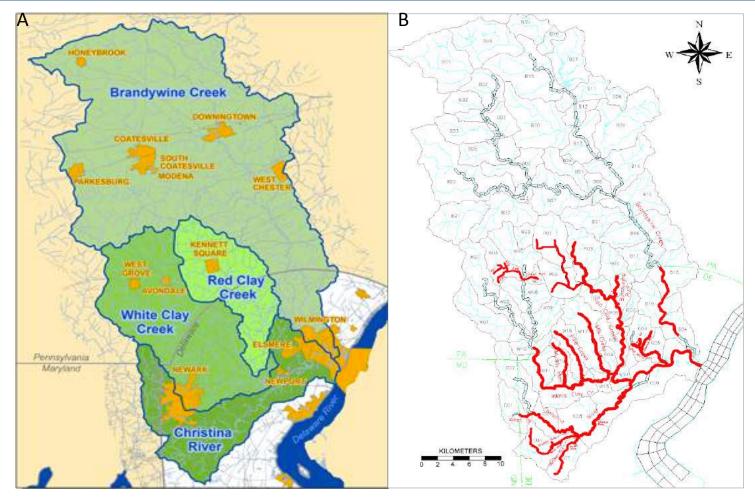
- Clean Water Act (CWA): "to restore and maintain the chemical, physical, and biological integrity of the Nation's waters"
- Impaired waters and TMDL (Total Maximum Daily Load) program
- Among all the TMDLs (including nutrients, sediments etc.), microbial contaminants (e.g. pathogenic bacteria) are ranked No. 1 causes for water quality degradation (U.S. EPA).

(39% rivers and streams, 13% of lakes, reservoirs and ponds;30% of assessed bays and estuaries)

Impaired waterways in US



Brandywine-Christina Basin



A: The Brandywine-Christina Basin includes Brandywine, White Clay, Red Clay and Christina river subwatersheds (adapted from Water Resource Agency, Univ. of Delaware). B: Stream segments impaired by bacteria (highlighted in red) by PA DEP and DNREC (adapted from U.S. EPA Brandywine-Christina Basin nutrients and bacterial TMDL).

Fecal Indicator Bacteria monitoring

- ✤ Total coliform → Fecal coliform → E. coli /Enterococcus
 - Public health agencies have used total coliforms and fecal coliforms as indicators since 1920s
 - non-fecal origin bacterial groups
 - coliforms can regrow in natural environments
 - Still being used in many states and agencies
 - *E. coli* and *Enterococcus*
 - More specific bacterial groups
 - Commonly used in these days
 - Recommended by US EPA (2012)

What bacteria should I monitor?

Depends on what you want to know; Consumption vs. recreation

• Health risk from recreational water contact:

Best indicators in freshwater: *E. coli* and *Enterococci*; for salt water, *Enterococci*

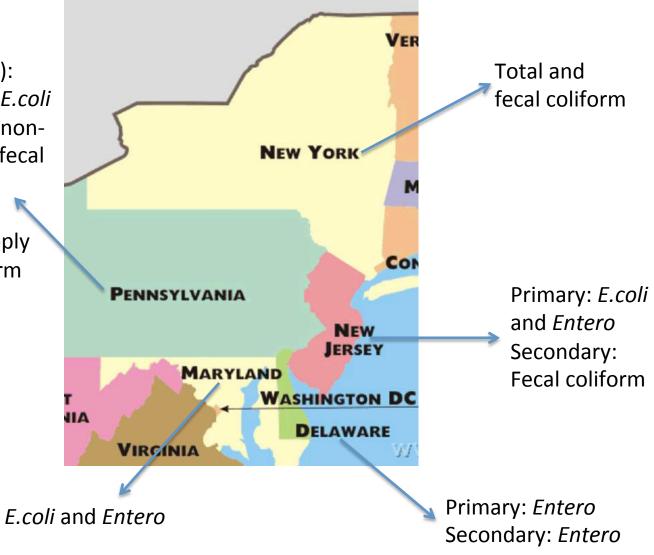
• Water supply or meets state water quality standards

Total and/or fecal coliform

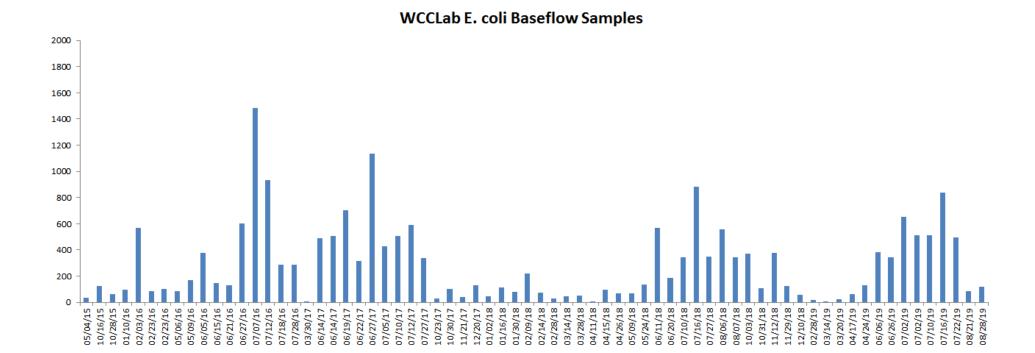
State Bacteria Monitoring

Water contact (WC): Swimming season- *E.coli* and fecal coliform; nonswimming season- fecal coliform.

Drinking Water supply (PWC): Total coliform

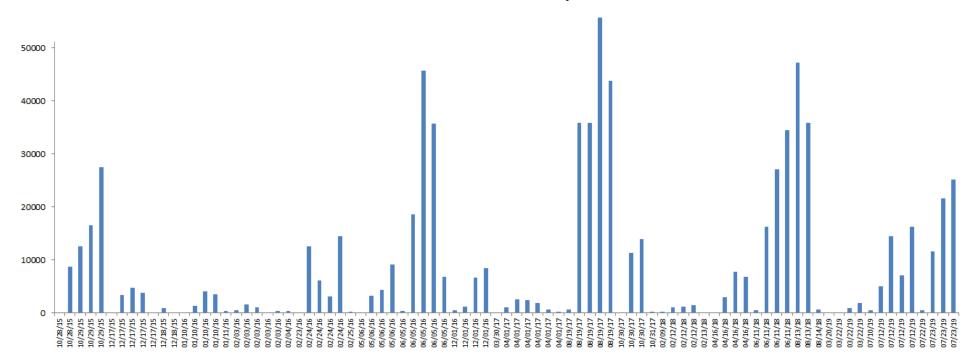


E. coli monitoring at WCC

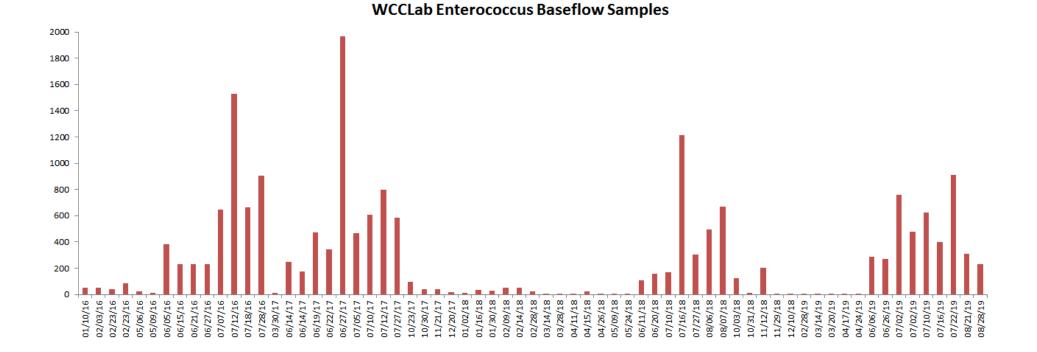


E. coli monitoring at WCC

WCCLab E. coli Storm Samples

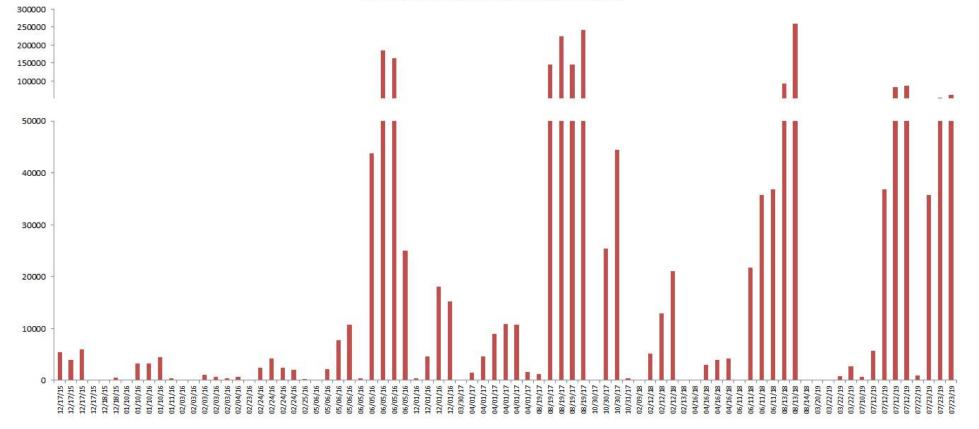


Enterococci monitoring at WCC



Enterococci monitoring at WCC

WCCLab Enterococcus Storm Samples



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Agriculture and urban







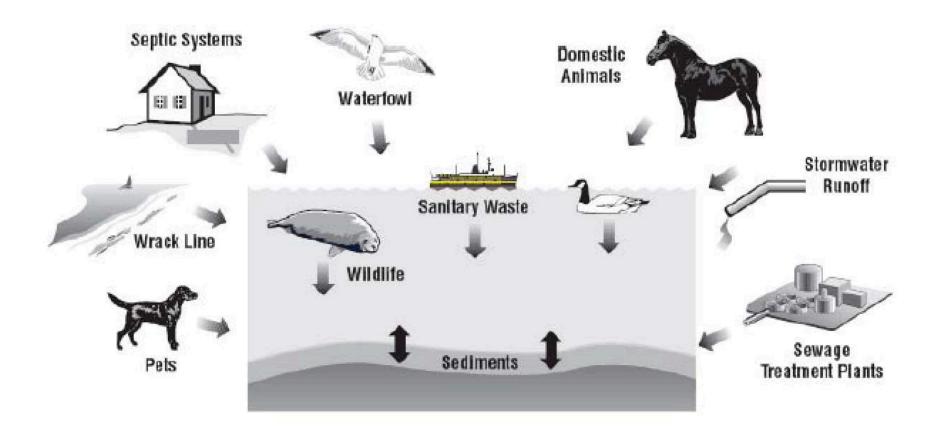


Wastewater treatment plant



Forested

Source of bacterial contaminants



Point and non-point sources

Source of bacterial contaminants

Sewage disposal systems: sewer breaks, sewer overflows, and sewer misconnections; on site septic systems

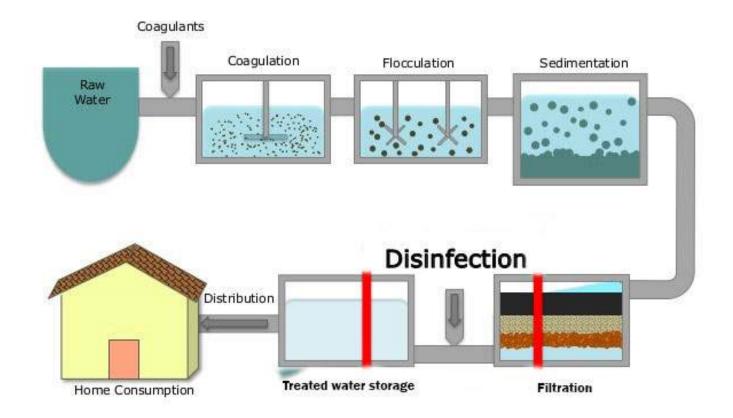
Agriculture: animal waste runoff, manure storage, vegetative buffer strips; livestock

Stormwater runoff: impervious surfaces, lacking catch basins and settling basins, inappropriate landscaping

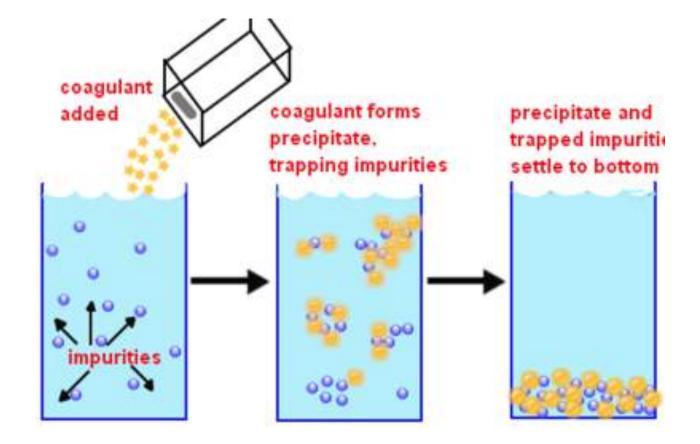
Wildlife: birds and small mammals; direct contact or watershed runoff; *Giardia, Cyrptosporidium, Salmonella, Camphylobacter, E. coli* etc.

Water treatment

Water Treatment Process



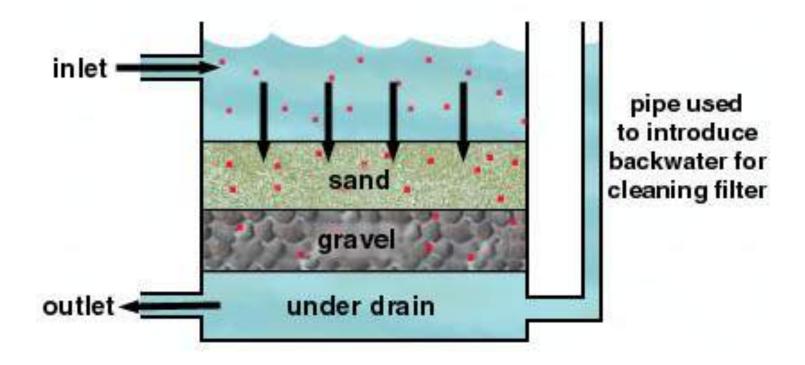
Water treatment



Coagulation

http://www.chemistry.wustl.edu/~edudev/LabTutorials/Water/PublicWaterSupply/PublicWaterSupply.html

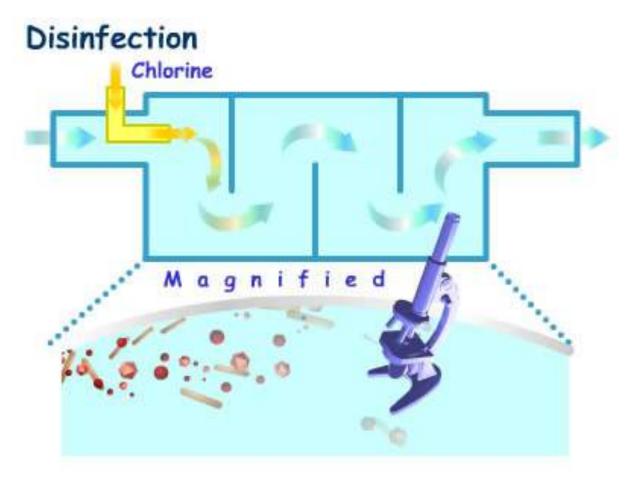
Water treatment



Filtration

http://www.chemistry.wustl.edu/~edudev/LabTutorials/Water/PublicWaterSupply/PublicWaterSupply.html

Water treatment



Disinfection

http://techalive.mtu.edu/meec/module03/Sources-SurfaceWater.htm

Prevention

Disinfection has proven effective and efficient against bacteria and enteric viruses, but protozoa such *Giardia* and especially *Cryptosporidium* are very resistant to chlorination alone!

The most important and cost effective protection for water suppliers is to **prevent** pathogen entry into source water.

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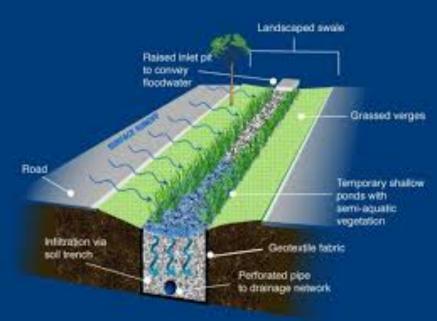
Remediation and restoration

Different approaches and BMPs Restoration efforts: Pasture management, runoff management, riparian protection, manure management etc.

- Buffer strips
- Constructed/storm water wetland
- Sand filters
- Retention/detention ponds
- Biofiltration

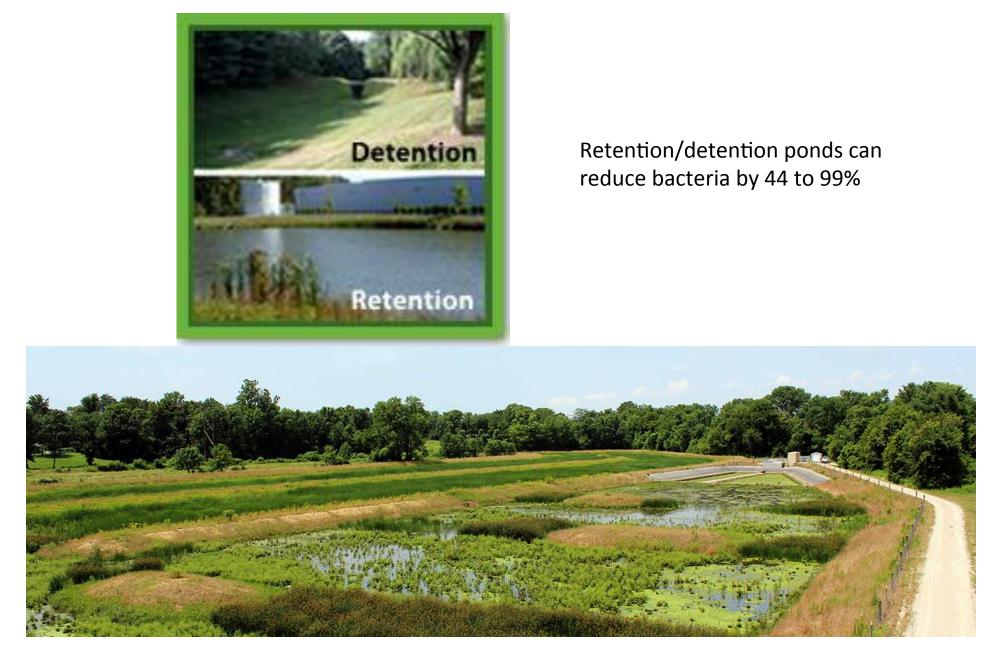


Buffers can reduce bacteria by 43 to 57%, especially in agricultural watersheds



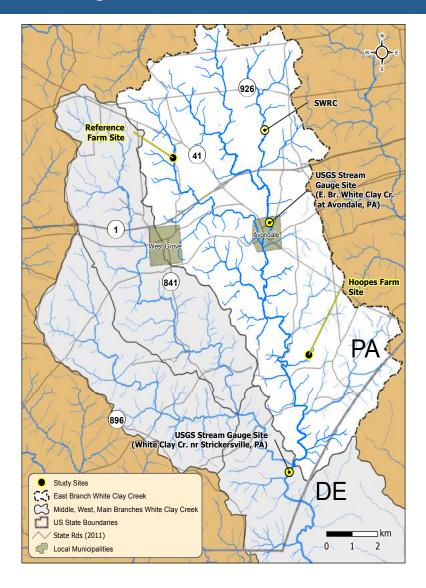
Biofiltration can reduce >99% of the microbes

(A. Boyer, DNREC, DE)



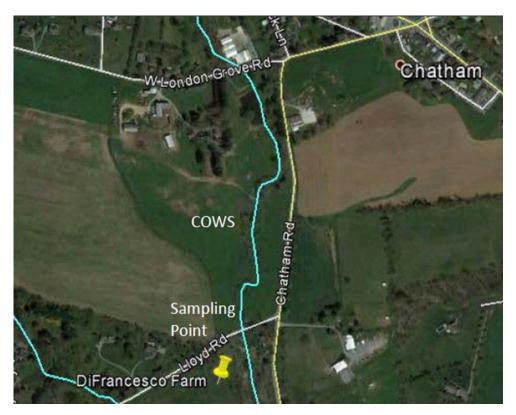
Constructed/storm water wetlands can reduce bacteria by 78 to 90%

Case study : United water/Suez



Case study : United water/Suez



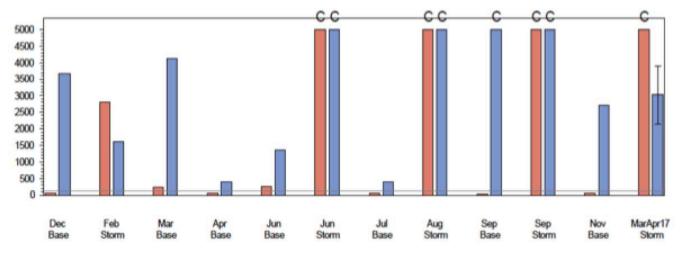


Reference site

BMP site

Keep livestock out of the streams!





Keep livestock out of the streams!



Fencing livestock out of streams is a highly effective method of reducing the amount of bacteria in surface waters

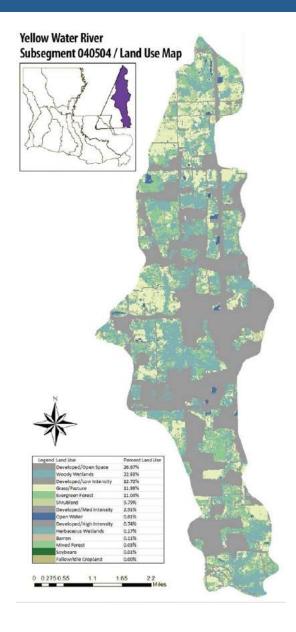
Case study: Yellow Water River

Yellow Water River in Louisiana

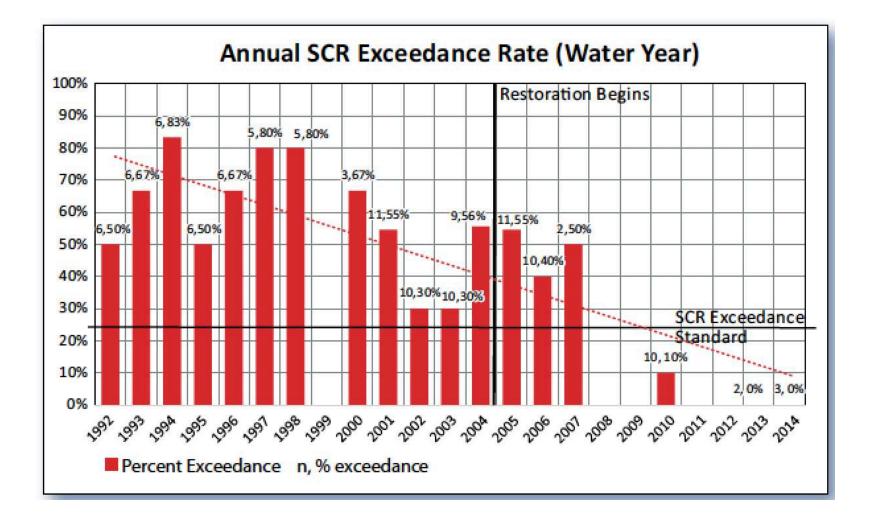
 Poor installation/maintenance on on-site treatment systems (septic system etc.)

Approaches

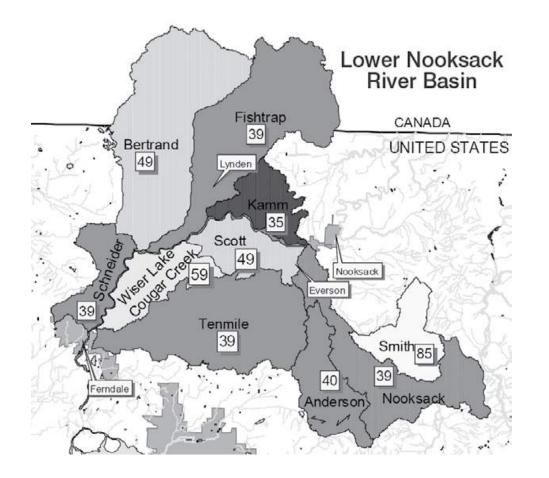
- Thorough inspections on waste water treatment plants and home waste systems
- Additional restoration activities including educational outreach, sewage inspections, and water quality monitoring etc.



Case study: Yellow Water River



Case study: Upper Fishtrap Creek

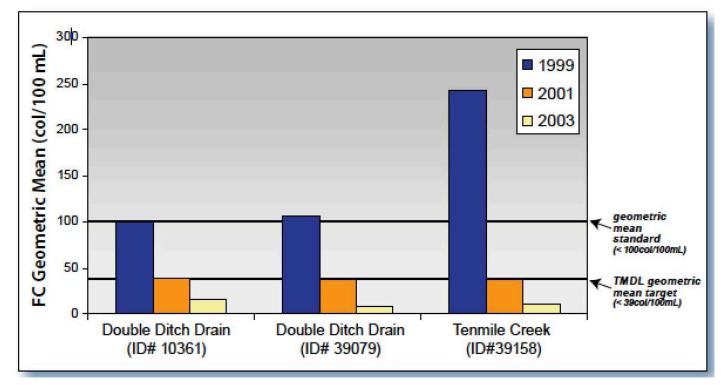


Lower Nooksack River in Washington

- Exceed 100 CFU/100ml
- FC impairments: state's CWA section 303(d) list

Case study: Upper Fishtrap Creek

- Nutrient management plans for all dairies
- Fence animals out
- Install hedgerows and filter srips
- On-site inspection and improvement on septic systems
- Farmers growing trees program



US EPA

Jinjun Kan, PhD Stroud Water Research Center

> jkan@stroudcenter.org 610 268 2153 ext 280



2012 EPA RWQC

A 30-day period geometric mean

Table 4. Recommended 2012 RWQC.

Criteria	36 per 1,000 p recre	ess Rate (NGI): rimary contact eators	-	Estimated Illness Rate (NGI) 32 per 1,000 primary contac recreators		
Elements	GM	nitude STV	-	GM	nitude STV	
Indicator	$(cfu/100 \text{ mL})^{a}$	$(cfu/100 \text{ mL})^{a}$	OR	$(cfu/100 \text{ mL})^{a}$	$(cfu/100 \text{ mL})^{a}$	
Enterococci – marine and fresh	35	130		30	110	
OR	h: il		1			
<i>E. coli</i> – fresh	126	410		100	320	

magnitude in any 30-day interval. There should not be greater than a ten percent excursion frequency of the selected STV magnitude in the same 30-day interval.

^a EPA recommends using EPA Method 1600 (U.S. EPA, 2002a) to measure culturable enterococci, or another equivalent method that measures culturable enterococci and using EPA Method 1603 (U.S. EPA, 2002b) to measure culturable *E. coli*, or any other equivalent method that measures culturable *E. coli*.

Beach Action Values

Table 5. Beach Action Values (BAVs).

	Estimated Illness Rate (NGI): 36 per 1,000 primary contact recreators		Estimated Illness Rate (NGI): 32 per 1,000 primary contact recreators
Indicator	BAV (Units per 100 mL)		BAV (Units per 100 mL)
Enterococci – culturable (fresh and marine) ^a	70 cfu		60 cfu
<i>E. coli</i> – culturable $(fresh)^{b}$	235 cfu	OR	190 cfu
Enterococcus spp. – qPCR (fresh and marine) ^c	1,000 cce		640 cce

^a Enterococci measured using EPA Method 1600 (U.S. EPA, 2002a), or another equivalent method that measures culturable enterococci.

^b *E. coli* measured using EPA Method 1603 (U.S. EPA, 2002b), or any other equivalent method that measures culturable *E. coli*.

^c EPA Enterococcus spp. Method 1611 for qPCR (U.S. EPA, 2012b). See section 5.2.

Pennsylvania

TABLE 3

Parameter Symbol Critical Use* Criteria ****** [(Fecal coliforms/ 100 ml)] (Escherichia coli/100 ml) -During the swimming season Bacteria Bacı WC (May 1 through September 30), the maximum [fecal coliform] E. coli level shall be a geometric mean of [200] 126 per 100 milliliters (ml) based on [a minimum of five] consecutive samples, each sample collected on different days, during a 30-day period. No more than 10% of the total samples taken during a 30-day period may exceed [400] 410 per 100 ml. (Fecal coliforms/ 100 ml)—For the remainder of the year, the maximum fecal coliform level shall be a geometric mean of 2,000 per 100 milliliters (ml) based on a minimum of five consecutive samples collected on different days during a 30-day period.] (Coliforms/100 ml)—Maximum of 5,000/100 ml as a monthly average value, no more PWS] [Bacz than this number in more than 20% of the samples collected during a month, nor more than 20,000/100 ml in more than 5% of the samples.

4.5.7 Bacterial Water Quality Criteria

Delaware

4.5.7.1 Primary and Secondary Contact Recreation Waters:

The following criteria shall apply:

Waterbody Type	Single-Sample Value (Enterococcus Colonies/100ml)	Geometric Mean (Enterococcu s Colonies/ 100ml)	
Primary Contact Recreation Fresh Waters	185	100	
Primary Contact Recreation Marine Waters	104	35	
Secondary Contact Recreation Fresh Waters	925	500	
Secondary Contact Recreation Marine Waters	520	175	

703.4 Water quality standards for coliforms.

Total and fecal coliform standards for specific classes are provided in this section.

New York

(a) Total coliforms (number per 100 ml).

Classes	Standard
AA	The monthly median value and more than 20 percent of the samples, from a minimum of five examinations, shall not exceed 50 and 240, respectively.
A, B, C, D, SB, SC, I, SD	The monthly median value and more than 20 percent of the samples, from a minimum of five examinations, shall not exceed 2,400 and 5,000, respectively.
SA	The median most probable number (MPN) value in any series of representative samples shall not be in excess of 70.
A-Special	The geometric mean, of not less than five samples, taken over not more than a 30-day period shall not exceed 1,000.
GA	The maximum allowable limit is 50.
(b) Fecal coliforms (number per 100 ml).	
A, B, C, D, SB, SC, I, SD	The monthly geometric mean, from a minimum of five examinations, shall not exceed 200.
A-Special	The geometric mean, of not less than five samples, taken over not more than a

30-day period shall not exceed 200.

(c) The total and fecal coliform standards for classes B, C, D, SB, SC and I shall be met during all periods:

(1) when disinfection is required for SPDES permitted discharges directly into, or affecting the best usage of, the water; or

(2) when the department determines it necessary to protect human health.

6 CRR-NY 703.4 Current through September 15, 2016

Maryland

A. Criteria for Class I Waters - Water Contact Recreation and Protection of Nontidal Warmwater Aquatic Life.

(1) Bacteriological.

(a) Table 1. Bacteria Indicator Criteria for Frequency of Use.

Steady State Geometric Mean Indicator Density		Single Sample Maximum Allowable Density				
Indicator Freshy	All Areas	Frequent Full Body Contact Recreation (Upper 75% CL)	Moderately Frequent Full Body Contact Recreation (Upper 82% CL)	Occasional Full Body Contact Recreation (Upper 90% CL)	Infrequent Full Body Contact Recreation (Upper 95% CL)	
(Either	Second Second Second					
Enterococci	33	61	78	107	151	
E. coli	126	235	298	410	576	
Marine	water					
Enterococci	35	104	158	275	500	

CL = confidence level

All numbers are counts per 100 milliliters

(b) In freshwater for E. coli, the following formula is used to calculate the upper 75 percent confidence interval for single sample maximum allowable density: antilog[(log 126) + 0.675 * log(SD)].

(c) In freshwater for enterococci, the following formula is used to calculate the upper 75 percent confidence interval for single sample maximum allowable density: antilog[($\log 33$) + 0.675 * $\log(SD)$], where $\log(SD)$ is the standard deviation of the log transformed E. coli or enterococci data. If the site data are insufficient to establish a log standard deviation, then 0.4 is used as the log standard deviation for both indicators. At the default log standard deviation, the values are 235 for E. coli and 61 for enterococci.

(d) In saltwater, for enterococci, the following formula is used to calculate the upper 75 percent confidence interval for single sample maximum allowable density: antilog[(log 35) + 0.675 * log(SD)], where log(SD) is the standard deviation of the log transformed enterococci data. If the site data are insufficient to establish a log standard deviation, then 0.7 is used as the log standard deviation. At the default log standard deviation, the value is 104.

New Jersey

- Bacterial quality (Counts/100 ml)
- Shellfish Harvesting: Bacterial Indicators shall not exceed, in all shellfish waters, the standard for approved shellfish waters as established by the National Shellfish Sanitation Program as set forth in its current manual of operations.
 - ii. Primary Contact Recreation:
 - Enterococci levels shall not exceed a geometric SE1 and SC mean of 35/100 ml, or a single sample maximum of 104/100 ml.
 - (2) E. Coli levels shall not exceed a geometric mean All FW2 of 126/100 ml or a single sample maximum of 235/100 ml.
 - iii. Secondary Contact Recreation:
 - Fecal coliform levels shall not exceed a geometric SE2 mean of 770/100 ml.
 - (2) Fecal coliform levels shall not exceed a geometric SE3 mean of 1500/100ml.